# 5-Selenization of salicylic acid derivatives yielded isoform-specific 5-lipoxygenase inhibitors†

Sun-Chol Yu,<sup>*a,b*</sup> Hartmut Kuhn,<sup>*a*</sup> Constantin-Gabriel Daniliuc,<sup>*c*</sup> Igor Ivanov,<sup>*a*</sup> Peter G. Jones<sup>*c*</sup> and Wolf-Walther du Mont<sup>*c*</sup>

Received 10th September 2009, Accepted 18th November 2009 First published as an Advance Article on the web 23rd December 2009 DOI: 10.1039/b918778b

Low molecular weight seleno-organic compounds exhibit glutathione peroxidase (GPx)-like activity; the well-known compound ebselen is being used in clinical trials as a stroke medication. Here, we describe the facile one-step synthesis of novel 5-selenized salicylic acid derivatives using selenium tetrachloride. The products were analyzed by spectroscopic studies including <sup>77</sup>Se-NMR and some were subjected to X-ray structure determination. Several products were identified as selective inhibitors of the pro-inflammatory 5-lipoxygenase (LOX) but had little effect on the catalytic activity of 12/15-LOX, which has been implicated in the synthesis of anti-inflammatory mediators. Such isoform-specificity (specificity coefficient >120) has not been reported before for any seleno-organic compound. In addition, synthesis products exhibited GPx-like activity, which was higher than that of ebselen for some derivatives.

### Introduction

GPx is an important constituent of the antioxidative defence system<sup>1</sup> and most mammalian GPx-isoforms bear a selenocysteine at the active site.<sup>2</sup> Many selenium-containing low molecular weight compounds also exhibit GPx activity in the presence of electron donors, and the chemistry of the catalytic reaction has been explored for ebselen<sup>3</sup> and other seleno-organic GPx mimetics.<sup>4,5</sup>

There are several ways to incorporate selenium into bioorganic molecules; selenization of aromatic rings<sup>6,7</sup> is a simple but effective principle. In 1938, 5-selenization of some salicylic acid esters using selenium tetrachloride (SeCl<sub>4</sub>) was attempted,<sup>8</sup> whereby selenium trihydroxides were initially suggested to be major products (Scheme 1, 1) that are nowadays regarded as seleninic acids (see Scheme 2).



Scheme 1 First 5-selenization of salicylic acid derivatives by SeCl<sub>4</sub>.

<sup>c</sup>Institute of Inorganic and Analytic Chemistry, Technical University of Braunschweig, Hagenring 30, 38106 Braunschweig, Germany. E-mail: p.jones@tu-bs.de; Tel: +49-531-3915307

<sup>†</sup> Electronic supplementary information (ESI) available: Spectral data for UV, IR, HR-MS (ESI) and NMR (<sup>1</sup>H, <sup>13</sup>C and <sup>77</sup>Se) of the synthesized compounds. CCDC 740213–740215. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/b918778b



Scheme 2 Synthetic route for 5-selenization of salicylic acid derivatives.

We have recently extended the synthetic scheme to a salicyloylglycine derivative (Scheme 2, **2a**).<sup>5</sup> This 5-selenization product exhibited a 4-fold higher GPx activity than ebselen,<sup>9</sup> the "gold standard" of seleno-organic compounds. Moreover, it inhibited plant and mammalian 12/15-LOXs at low concentration.<sup>5</sup>

LOXs form a heterogeneous family of lipid-peroxidizing enzymes, which have also been implicated in inflammatory<sup>10</sup> and hyperproliferative<sup>11</sup> diseases. In humans, there are 6 different LOX isoforms originating from separate genes.<sup>12</sup> The isoform 5-LOX is involved in the biosynthesis of leukotrienes, which

<sup>&</sup>lt;sup>a</sup>Institute of Biochemistry, University Medicine Berlin-Charité, Monbijoustrasse 2, 10117 Berlin, Germany. E-mail: hartmut.kuehn@charite.de; Fax: +49-30-450528905; Tel: +49-30-450528040

<sup>&</sup>lt;sup>b</sup>Department of Pharmacy, Pyongyang University of Medicine, Kangandong, Songyo district, Pyongyang, DPR of Korea. E-mail: sun-chol.yu@ charite.de; Tel: +49-30-450528018

constitute powerful pro-inflammatory mediators,<sup>13</sup> and the 5-LOX inhibitor zileuton has been approved for clinical use.<sup>14</sup> In contrast, anti-inflammatory activities<sup>15</sup> have been reported for 12/15-LOX and epidermal-type LOX has been implicated in skin development.<sup>16</sup> Because of this functional multiplicity, the search for isoform-specific LOX inhibitors is a challenging task in current pharmacological research. Owing to their redox characteristics, several seleno-organic compounds have been identified as LOX inhibitors but isoform-specificity for human isozymes has been not reported.

In this context, we synthesized novel 5-selenized salicylic acid derivatives and tested their GPx-activity as well as their isoform-dependent inhibition of two major LOX isoforms (5-LOX and 12/15-LOX).

#### **Results and discussion**

#### Synthesis

5-Selenization was initiated by reacting salicylic acid derivatives with  $SeCl_4$  in tetrahydrofuran (THF), which is known to be a good solvent for  $SeCl_4$  (Scheme 2).

Under mild reaction conditions (RT, 1 h) yellow 5-selenium trichlorides were formed,<sup>8</sup> which were further converted in water to white 5-seleninic acids (3) or to the corresponding anhydrides (2). Here it should be pointed out that we could not observe any formation of 5-selenium trihydroxides (1), which had previously been analyzed without using the benefits of modern techniques such as NMR, HR-MS or X-ray crystallography.<sup>8</sup> The crystal data on ethyl ester (3d) reveals that a seleninic acid is produced in the reaction. It is interesting, moreover, that salicylic acid amides bearing a free terminal carboxylic group were converted to 5-seleninic acid anhydrides (Scheme 2, 2a and 2b), whereas those lacking this structural element yielded 5-seleninic acids (Scheme 2, 3c–e). This phenomenon will be discussed in the next subsection.

When the reaction mixture (Scheme 2) was refluxed for 1 h or stirred for one day at RT, the 5-selenium trichloride decomposed to form a mixture of yellow diselenide (4) and white monoselenide (5), which were separated by flash chromatography on silica gel.

#### X-Ray crystallography<sup>17</sup>

All molecules display an intramolecular hydrogen bond from the OH group to the neighbouring carbonyl function (Fig. 1, 3 and 5; ellipsoids represent 50% probability levels). The packing involves a mixture of classical hydrogen bonds and Se  $\cdots$  LO contacts (Fig. 2, 4 and 6); for the latter we have set an arbitrary limit of 3.2 Å, above which the contacts are not considered (the next longest are *ca*. 3.4 Å).

Compound **3c** has a complex three-dimensional packing involving four independent contacts: three hydrogen bonds O2–H02···O1<sup>i</sup> 2.636(2) Å, N1–H1A···O3<sup>ii</sup> 2.892(2) Å, N1–H1B···O1<sup>iii</sup> 2.932(2) Å, and one Se···O3(iv) contact of 3.166(2) Å (symmetry operations are  $i = (\frac{1}{2} + x, \frac{3}{2} - y, 2 - z)$ ,  $ii = (-x, \frac{1}{2} + y, \frac{3}{2} - z)$ ,  $iii = (-x, -\frac{1}{2} + y, \frac{3}{2} - z)$ ,  $iv = (1 - x, \frac{1}{2} + y, \frac{3}{2} - z)$ ). Fig. 2 shows a two-dimensional ribbon substructure involving the NH<sub>2</sub> groups. The third H bond connects this ribbon to a perpendicularly oriented neighbour, whereas the Se···O3 contact links to a parallel layer.



**Fig. 1** Structure of compound **3c** in the crystal. Bond lengths (Å) and angles (°) at selenium: Se–O1 1.659(1), Se–O2 1.756(1), Se–C1 1.921(2); O1–Se–O2 104.6(1), O1–Se–C1 101.1(1), O2–Se–C1 93.3(1).



Fig. 2 Packing diagram of compound 3c.

For compound **3d**, the hydrogen bonds O2–H02···O1<sup>i</sup> 2.547(2) Å and O5–H05···O4 2.655(2) Å combine with Se ···O1<sup>ii</sup> 3.048(1) Å contact to produce a layer structure parallel to the *xy* plane (Fig. 4), (symmetric operations  $i = (-x, -\frac{1}{2} + y, \frac{1}{2} - z)$ , ii = (-x,  $-\frac{1}{2} + y, \frac{1}{2} - z)$ ); the aromatic rings lie approximately perpendicular to this plane.

Crystallization of compound **3e** from anhydrous methanol led to single crystals of its seleninic methyl ester derivative **6**. For compound **6**, one hydrogen bond O3–H03···O1<sup>i</sup> 2.575(2) Å combines with Se···O1<sup>ii</sup> 2.946(2) Å and Se···O4<sup>iii</sup> 3.182(1) Å contacts to form a layer structure parallel to the *yz* plane (Fig. 6) (symmetry operations  $i = (1 - x, \frac{1}{2} + y, \frac{3}{2} - z)$ , ii = (1 - x, 1 - y, 2 - z),  $iii = (1 - x, -\frac{1}{2} + y, \frac{3}{2} - z)$ ).

The crystallographic data are shown in Table 1.

The methyl esterification mentioned above was also observed in the HR-MS spectrum, in which MeOH was used as a solvent. As seen in Fig. 7, the methyl ester was the main peak (264.96031). NMR and elemental analysis of **3e** proves the possibility of its rapid esterification in MeOH.

This phenomenon indirectly indicates that seleninic acid derivatives with a carboxyl group might have a potential tendency toward esterification and, therefore, salicylic acid amides carrying a free



**Fig. 3** Structure of compound **3d** in the crystal. Bond lengths (Å) and angles (°) at selenium: Se–O1 1.666(1), Se–O2 1.744(1), Se–C1 1.934(2); O1–Se–O2 105.8(1), O1–Se–C1 98.5(1), O2–Se–C1 99.0(1).



Fig. 4 Packing diagram of compound 3d.

terminal carboxylic group such as **2a** and **2b** could be produced through self-dehydration (so-called self-esterification).

#### **GPx-like** activity

The GPx-like activity of the synthesized compounds was assayed in coupled reductase model<sup>5</sup> with the use of glutathione and *t*-BuOOH as substrates. As expected from the reaction mechanism catalyzed by compound 2a,<sup>5</sup> 5-seleninic acid and diselenide derivatives (**3c–e**, **4e**) also exhibited a high GPx-like activity (Table 2).

From the comparison of the 5-seleninic acid (3e) with the diselenide type (4e) it can be seen that if normalized to selenium content, the former is more effective than the latter in GPx-like activity (ratio 2.8:2.1), but that these two moieties are certain to be major active components<sup>5</sup> in the catalytic cycle of 5-selenized salicylic acid derivatives. The inactivity of monoselenide



**Fig. 5** Structure of compound **6** in the crystal. Bond lengths (Å) and angles (°) at selenium: Se–O1 1.666(1), Se–O2 1.774(2), Se–C1 1.927(2); O1–Se–O2 104.8(1), O1–Se–C1 100.6(1), O2–Se–C1 97.0(1).



Fig. 6 Packing diagram of compound 6.



**5e** was predicted from previous work.<sup>5</sup> The lack of the activity for compound **2b** suggests that a long aliphatic chain may inhibit the catalytic function, which could bring about a new concept in understanding the mechanism of selenium-containing GPx mimics.

## Table 1 Crystallographic data for compounds 3c, 3d and 6

| Compound   | 3c   | 3d   | 6   |
|--|--|--|---|
| Formula  | C <sub>7</sub> H <sub>7</sub> NO <sub>4</sub> Se | C <sub>9</sub> H <sub>10</sub> O <sub>5</sub> Se | C <sub>8</sub> H <sub>8</sub> O <sub>5</sub> Se |
| $M_{\rm r}$  | 248.10   | 277.13   | 263.11  |
| Habit  | Colourless needle                                | Colourless needle                                | Colourless prism                                |
| Cryst. size/mm                                       | $0.16 \times 0.12 \times 0.10$                   | $0.2 \times 0.1 \times 0.1$                      | $0.12 \times 0.11 \times 0.09$                  |
| Crystal system                                       | Orthorhombic                                     | Monoclinic                                       | Monoclinic                                      |
| Space group  | $P2_{1}2_{1}2_{1}$                               | $P2_{1}/c$                                       | $P2_{1}/c$                                      |
| Cell constants:                                      |  |  |   |
| a/Å  | 4.9234(2)  | 11.9653(4)                                       | 5.1610(2)                                       |
| b/Å  | 9.5712(2)  | 5.1029(2)  | 17.4964(8)                                      |
| c/Å  | 17.3190(6)                                       | 17.4167(8)                                       | 10.2112(6)                                      |
| $\alpha$ (°)   | 90   | 90   | 90  |
| $\hat{\boldsymbol{\beta}}(\hat{\boldsymbol{\circ}})$ | 90   | 110.947(4)                                       | 90.578(4)                                       |
| $\gamma$ (°)   | 90   | 90   | 90  |
| $V/Å^3$  | 816.12   | 993.14   | 922.01  |
| Z  | 4  | 4  | 4   |
| $D_{\rm x}$ (Mg m <sup>-3</sup> )                    | 2.019  | 1.853  | 1.895   |
| $\mu/mm^{-1}$  | 4.6  | 3.8  | 5.5   |
| F(000)   | 488  | 552  | 520   |
| T/°C   | -173   | -173   | -173  |
| Radiation, $\lambda/Å$                               | Μο-Κα, 0.71073                                   | Μο-Κα, 0.71073                                   | Cu-Ka, 1.54184                                  |
| $2\theta_{\rm max}$                                  | 60   | 58   | 151   |
| Refl. measured                                       | 18 703   | 22715  | 16 694  |
| Refl. indep.   | 2338   | 2658   | 1898  |
| $R_{\rm int}$  | 0.038  | 0.037  | 0.035   |
| Parameters   | 134  | 154  | 136   |
| Restraints   | 2  | 16   | 0   |
| $WR(F^2, all refl.)$                                 | 0.035  | 0.046  | 0.062   |
| $R(F, >4\sigma(F))$                                  | 0.020  | 0.021  | 0.025   |
| S  | 0.93   | 0.93   | 1.12  |
| max. $\Delta \rho / e \text{ Å}^{-3}$                | 0.56   | 0.54   | 0.38  |

**Table 2** The rate of NADPH oxidation  $(v_0)$  in presence of synthesized compounds (10  $\mu$ M) and their GPx-like activity relative to ebselen (n = 3)

| Compound       | $\upsilon_0/\mu M \min^{-1}$ | Relative activity |
|----------------|------------------------------|-------------------|
| Ebselen        | $5.85 \pm 0.76$              | 1                 |
| 2b             | Not detectable               | 0                 |
| 3c             | $12.05 \pm 1.82$             | 2.1               |
| 3d             | $5.21 \pm 0.28$              | 0.9               |
| 3e             | $16.40 \pm 1.02$             | 2.8               |
| 4e             | $24.63 \pm 2.65$             | 4.2               |
| 5e             | Not detectable               | 0                 |
| Salicylic acid | Not detectable               | 0                 |

The high GPx-like activity of synthesized 5-seleninic acids is consistent with the more recent suggestion<sup>3</sup> that the seleninic acid moiety is also one of the intermediates in the GPx-like activity cycle of ebselen.

# LOX inhibition

Rabbit reticulocyte 12/15-LOX inhibition. As shown in Table 3, 5-seleninic acid derivatives (2b, 3c-e) and monoselenide 5e did not inhibit 12/15-LOX effectively. In contrast, the diselenide 4e inhibited the enzyme with an IC<sub>50</sub> of about 5  $\mu$ M (Table 3).

# **Recombinant human 5-LOX inhibition**

Almost all of the 5-selenized compounds inhibited recombinant human 5-LOX at lower micromolar concentrations (Table 4).

Among them, compound 3d, with an  $IC_{50}$  of 0.66  $\mu$ M, was most effective and over 200 times more specific to 5-LOX than to 12/15-

**Table 3** 12/15-LOX inhibition of synthesized compounds (n = 3)

| Compound | Inhibition at 100 $\mu$ M (%) | $IC_{50}/\mu M$ |
|----------|-------------------------------|-----------------|
| 2b       | $6.07 \pm 1.85$               |                 |
| 3c       | $16.96 \pm 3.26$              |                 |
| 3d       | $45.29 \pm 2.44$              | 146             |
| 3e       | $1.19 \pm 2.69$               |                 |
| 4e       | $96.58 \pm 0.77$              | 5.36            |
| 5e       | $7.31 \pm 1.62$               |                 |

Table 4 5-LOX inhibition of synthesized compounds (n = 3) and their relative specificity

| Compound | $IC_{50}/\mu M$ | Ratio (12/15-LOX/5-LOX) |
|----------|-----------------|-------------------------|
| Ebselen  | 1.09            | 0.51                    |
| 2a       | 1.01            | 7.82                    |
| 2b       | 0.79            | >127                    |
| 3c       | 0.80            | >125                    |
| 3d       | 0.66            | 221                     |
| 3e       | 2.37            | >42                     |
| 4e       | 2.09            | 2.56                    |
| 5e       | 4.94            | >20                     |

LOX. Because the specificity ratios  $(IC_{50}(12/15-LOX)/IC_{50}(5-10))$ LOX)) of 2b, 3c, 3e and 5e (Table 4) are based on 100 µM maximal test concentration for 12/15-LOX, the real values could be even higher than that of 3d (Fig. 8). The non-specific property of 4e might provide a chance to develop stronger general LOX inhibitors related to diselenides.

Zileuton (not a seleno-organic compound), the only 5-LOX inhibitor currently used in clinics, is less powerful (IC<sub>50</sub> of  $3.8 \,\mu$ M)



**Fig. 8** Comparison of **2b** ( $\blacksquare$ ), **3c** ( $\blacklozenge$ ) and **3d** ( $\blacktriangle$ ) with ebselen ( $\bigtriangledown$ ) in the selectivity between 12/15 LOX (---) and 5- LOX ( $\frown$ ) inhibition.

than compound **3d** in the model but exhibits a similar degree of isoform-specificity.<sup>18</sup> Although selenium-containing 5-LOX inhibitors (7–10) with similar or even lower IC<sub>50</sub> have been reported before in different models (Fig. 9),<sup>19,20</sup> their isoform-specificity has not been tested yet.



Fig. 9 Some known selenium-containing 5-LOX inhibitors.

Salicylic acid has recently been tested for the inhibition of several LOXs, where its  $IC_{50}$  for 12/15 and 5-LOXs was 49  $\mu$ M and 168  $\mu$ M, respectively,<sup>21</sup> and the specificity coefficient was, therefore, very poor (0.29).

These results indicate that 5-selenization might become a significant method for enhancing the specific inhibition of 5-LOX by salicylic acid derivatives.

The mechanism of 5-LOX inhibition of the seleno-organic compounds has not been explored in detail. However, previous investigations into the mode of action of ebselen<sup>22</sup> suggested that seleno-organic compounds may counteract LOX activation. In their silent form, LOXs contain ferrous non-heme iron, which is oxidized by hydroperoxide activators to its ferric form. We found that the seleno-organic compounds synthesized in this study are capable of reducing ferric iron (Fig. 10) and thus, there is the possibility that those compounds reduce the active ferric LOX to its catalytically silent ferrous form.

A necessary precondition for this mechanism is that the inhibitor is capable of penetrating the active site of the enzymes. The crystal structures of various LOX isoforms<sup>23</sup> suggested that the active site of LOX constitutes a deep hydrophobic U-shaped



Fig. 10 Iron-reducing activity of synthesized compounds.

pocket, which buries the catalytic iron. If an inhibitor is sufficiently small and hydrophobic it can penetrate the active site to interact with the iron. Unfortunately, the crystal structure of human 5-LOX has not been solved so far. However, structural modeling on the basis of the X-ray coordinates of rabbit 12/15-LOX suggested that the active site of human 5-LOX is larger than that of 12/15-LOX and mutagenesis studies on human 5-LOX support this hypothesis,<sup>24</sup> but non-relationship between LOX inhibitory activity and iron-reducing activity of the tested compounds would suggest that these structural differences in active sites of the enzymes might not explain the isoform-specific inhibitory effect of them.

Instead, it could be expected that 5-seleninic acid derivatives may contribute to interaction with the active site of 5-LOX through its property of esterification or dehydration mentioned above, as aspirin (acetyl salicylic acid) irreversibly inhibits cyclooxygenase by acetylation of the active site residue.

#### Conclusions

Most of the novel 5-selenized salicylic acid derivatives synthesized in this study exhibit a higher GPx-like activity than ebselen. In addition, all compounds inhibit the recombinant human 5-LOX at lower micromolar concentrations and the seleninic acid derivatives (**3c-e**) show a remarkable LOX isoform-specificity. More work is needed to explore the underlying mechanistic details.

#### Experimental

#### Materials and methods

Commercial reagents salicyloylglycine,  $\varepsilon$ -aminocaproic acid, ethyl salicylate, salicylic acid and selenium tetrachloride were purchased from Merck and Sigma-Aldrich. All solvents were of extra pure grade from Roth (Germany). IR spectra were recorded on a Nicolet Avatar 360 FT-IR. Kinetic measurements were performed on a Shimadzu UV-2102 spectrophotometer. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Brucker Biospin AV 400 (<sup>1</sup>H: 400 MHz; <sup>13</sup>C: 100.5 MHz) by using d<sub>6</sub>-DMSO as a solvent. Chemical shift values ( $\delta$ ) are reported in ppm downfield from TMS ( $\delta$  = 0.0 ppm) as internal standard. <sup>77</sup>Se NMR spectra were recorded by using a Brucker Avance II 300 instrument (57.20 MHz) in d<sub>6</sub>-DMSO (dimethylselenide as external standard). HR-MS recording was carried out on a Finnigan Thermo Scientific LTQ OrbitrapXL (ESI) in MeOH. Elemental analysis was carried out on HEKAtech EURO EA, vario EL.

# General procedure for 5-selenization of salicylic acid derivatives by selenium tetrachloride

Selenium tetrachloride 1.107 g (5 mmol) was added to a solution of salicylic acid derivatives (5 mmol) in 10 ml of THF and the mixture was stirred for 1 h at room temperature. Then, 100 ml of water was added to the reaction mixture and the sample was stirred at room temperature for an additional 24 h. The white precipitate was filtered, washed with  $3 \times 30$  ml of THF and dried in air at 30 °C. The product was purified by flash chromatography (*i*PrOH–NH<sub>4</sub>OH–H<sub>2</sub>O; 8:1:1). If the reaction runs under reflux for 1 h and follows the same procedure as above, one can obtain a mixture of monoselenide and diselenide as the main products, which are isolated by the same condition of flash chromatography. 6-[(2-Hydroxybenzoyl)amino]hexanoic acid as a starting material for synthesis of 2b was synthesized by reaction of ethyl salicylate with  $\varepsilon$ -aminocaproic acid in 25% sodium methylate solution MeOH and acidification with dilute acid. 2-[[5-[3-(carboxymethylcarbamoyl)-4-hydroxysulfuric phenyl]seleninyloxyseleninyl-2-hydroxy-benzoyl]amino]acetic acid (2a) was from the previous work.

**6-[(2-Hydroxybenzoyl)amino]hexanoic acid.** Yield: 76%, mp: 110–112 °C (lit.,<sup>25</sup> 107–108 °C), Found: C, 62.21; H, 6.51; N, 5.23. Calc. for  $C_{13}H_{17}NO_4$ : C, 62.14; H, 6.82; N, 5.57%,  $\delta_H$  1.31 (2 H, m, CH<sub>2</sub>), 1.53 (4 H, m, CH<sub>2</sub>), 2.21 (2 H, t, CH<sub>2</sub>), 3.28 (2 H, m, CH<sub>2</sub>), 6.90 (2 H, m, Ar–H), 7.39 (H, t, Ar–H), 7.85 (H, dd, Ar–H), 8.81 (H, t, NH), 12.00 (H, s, COOH), 12.72 (H, s, OH),  $\delta_C$  24.21 (CH<sub>2</sub>), 25.98 (CH<sub>2</sub>), 28.58 (CH<sub>2</sub>), 33.57 (CH<sub>2</sub>), 38.77 (CH<sub>2</sub>), 115.04 (C-1), 117.40 (C-3), 118.46 (C-6), 127.48 (C-4), 133.62 (C-5), 160.26 (C-2), 169.00 (C=O), 174.45 (COOH), *m/z* (ESI) 252.1232 (M<sup>+</sup>. C<sub>13</sub>H<sub>17</sub>NO<sub>4</sub> requires 252.1230).

**6-[[2-Hydroxy-5-[4-hydroxy-3-](6-hydroxy-6-oxo-hexyl)carbamoyl]phenyl]seleninyloxyseleninyl-benzoyl]amino]hexanoic acid (2b).** Yield: 63%, mp: 126–128 °C, Found: C, 44.24; H, 4.85; N, 3.88. Calc. for C<sub>26</sub>H<sub>32</sub>N2O<sub>11</sub>Se<sub>2</sub>: C, 44.20; H, 4.57; N, 3.97%,  $\delta_{\rm H}$ 1.31 (2 H, m, CH<sub>2</sub>), 1.53 (4 H, m, CH<sub>2</sub>), 2.21 (2 H, t, CH<sub>2</sub>), 3.29 (2 H, m, CH<sub>2</sub>), 7.10 (H, d, Ar–H), 7.82 (H, dd, Ar–H), 8.30 (H, d, Ar–H), 9.04 (H, t, NH), 13.16 (H, s, COOH),  $\delta_{\rm C}$  24.20 (CH<sub>2</sub>), 25.98 (CH<sub>2</sub>), 28.51 (CH<sub>2</sub>), 33.57 (CH<sub>2</sub>), 38.96 (CH<sub>2</sub>), 115.74 (C-1), 118.04 (C-3), 126.31 (C-6), 130.87 (C-4), 138.59 (C-5), 162.52 (C-2), 167.83 (C=O), 174.45 (COOH),  $\delta_{\rm Se}$  1177, *m/z* (ESI) 709.0409 (M<sup>+</sup>. C<sub>26</sub>H<sub>32</sub>N<sub>2</sub>O<sub>11</sub><sup>80</sup>Se<sub>2</sub> requires 709.0406).

**3-Carbamoyl-4-hydroxy-benzeneseleninic** acid (3c). Yield: 63%, mp: 154–156 °C, Found: C, 34.16; H, 2.86; N, 5.65. Calc. for C<sub>7</sub>H<sub>7</sub>NO<sub>4</sub>Se: C, 33.89; H, 2.84; N, 5.65%,  $\delta_{\rm H}$  7.11 (H, d, Ar–H), 7.83 (H, dd, Ar–H), 8.06 (H, s, Ar–H), 8.32 (H, d, NH), 8.60 (H, s, NH), 13.40 (H, s, OH),  $\delta_{\rm C}$  115.01 (C-1), 118.12 (C-3), 126.99 (C-6), 131.17 (C-4), 138.45 (C-5), 163.38 (C-2), 170.81 (C=O),  $\delta_{\rm se}$ 1177, *m*/*z* (ESI) 247.9474 (M<sup>-</sup>. C<sub>7</sub>H<sub>7</sub>NO<sub>4</sub><sup>80</sup>Se requires 247.9468).

**3-Ethoxycarbonyl-4-hydroxy-benzeneseleninic acid (3d).** Yield: 17%, mp: 122–123 °C, Found: C, 39.07; H, 3.45. Calc. for C<sub>9</sub>H<sub>10</sub>O<sub>5</sub>Se: C, 39.01; H, 3.64%,  $\delta_{\rm H}$  1.36 (3 H, t, CH<sub>3</sub>), 4.30 (2 H, dd, CH<sub>2</sub>), 7.16 (H, d, Ar–H), 7.93 (H, dd, Ar–H), 8.23 (H, d, Ar–H),  $\delta_{\rm C}$  14.01 (CH<sub>3</sub>), 61.66 (CH<sub>2</sub>), 113.75 (C-1), 118.08 (C-3), 128.12 (C-6), 132.89 (C-4), 139.37 (C-5), 162.24 (C-2), 167.88 (C=O),  $\delta_{\rm sc}$  1176, *m/z* (ESI) 276.96209 (M<sup>-</sup>. C<sub>9</sub>H<sub>10</sub>O<sub>5</sub><sup>80</sup>Se requires 276.96207).

**2-Hydroxy-5-selenino-benzoic acid (3e).** Yield: 15%, mp: 140– 142 °C (decomp.), Found: C, 33.46; H, 2.72. Calc. for  $C_7H_6O_3Se$ : C, 33.75; H, 2.43%,  $\delta_H$  7.12 (H, d, Ar–H), 7.90 (H, dd, Ar–H), 8.23 (H, d, Ar–H),  $\delta_C$  113.48 (C-1), 117.84 (C-3), 128.44 (C-6), 132.94 (C-4), 139.06 (C-5), 163.40 (C-2), 171.12 (C=O),  $\delta_{Se}$  1175, m/z (ESI) 248.93021 (M<sup>-</sup>.  $C_7H_6O_5^{80}Se$  requires 248.93077).

**5-(3-Carboxy-4-hydroxy-phenyl)diselanyl-2-hydroxy-benzoic acid (4e).** Yield: 13%, mp: 275 °C (decomp.) (lit.,<sup>26</sup> 231–235 °C), Found: C, 38.86; H, 2.23. Calc. for  $C_{14}H_{10}O_6Se_2$ : C, 38.91; H, 2.33%,  $\delta_H$  6.81 (H, d, Ar–H), 7.54 (H, dd, Ar–H), 7.92 (H, d, Ar–H),  $\delta_C$  116.37 (C-1), 117.74 (C-3), 118.14 (C-5), 135.54 (C-6), 139.40 (C-4), 162.42 (C-2), 170.84 (C=O),  $\delta_{Se}$  502, *m/z* (ESI) 432.87284 (M<sup>-</sup>.  $C_{14}H_{10}O_6^{80}Se_2$  requires 432.87350).

**5-(3-Carboxy-4-hydroxy-phenyl)selanyl-2-hydroxy-benzoic acid** (**5e**). Yield: 7%, mp: 262 °C (decomp.) (lit.,<sup>27</sup> 272 °C), Found: C, 47.48; H, 2.82. Calc. for  $C_{14}H_{10}O_6Se: C, 47.61; H, 2.85\%, \delta_H$  6.93 (H, d, Ar–H), 7.58 (H, dd, Ar–H), 7.85 (H, d, Ar–H),  $\delta_C$  114.35 (C-1), 118.77 (C-3), 119.25 (C-5), 134.77 (C-6), 140.26 (C-4), 160.75 (C-2), 170.90 (C=O),  $\delta_{Se}$  397, *m*/*z* (ESI) 352.95614 (M<sup>-</sup>.  $C_{14}H_{10}O_6^{80}Se$  requires 352.95698).

#### X-Ray structure determinations

**Data collection and reduction.** Crystals were mounted in inert oil on glass fibres and transferred to the cold gas stream of the diffractometer (**3c**, **3d**: Oxford Diffraction Xcalibur E with monochromated Mo-K $\alpha$  radiation; **6**: Oxford Diffraction Nova A with mirror-focussed Cu-K $\alpha$  radiation).

**Structure refinement.** The structures were refined anisotropically against  $F^{2,28}$  Hydrogen atoms of hydroxy and NH<sub>2</sub> groups were refined freely but with X–H distance restraints; other H atoms were included with a riding model or rigid methyl groups. For compound **3c**, the Flack parameter was refined to 0.002(8). For compound **3d**, the methoxy group was disordered over two positions (minor site occupation 19%), which were subjected to similarity restraints.

#### GPx-like acitivity5

The catalytic reaction was run at 37 °C in a 1 ml reaction mixture consisting of 100 mM Tris-HCl buffer (pH 7.4) containing 5 mM EDTA and 0.1% Triton X-100, 3 mM GSH, 0.2 mM NADPH, 1 U of glutathione reductase and  $10 \,\mu M$  of the test compound (DMSO solution). The assay sample was equilibrated for 10 min in the absence of peroxide substrate and GPx-like reaction was initiated by addition of 0.5 mM tert-butyl hydroperoxide (tBuOOH). The time-dependent decrease in absorbance at 340 nm (10-40 s after addition of tBuOOH) was recorded. During the GPx-like reaction, reduced GSH is oxidized and the resulting disulfide (GSSG) is back-reduced by the glutathione reductase reaction consuming stoichiometric amounts of NADPH. Since NADPH exhibits a local absorbance maximum  $(A_{cat})$  at 340 nm but its oxidized counterpart (NADP) does not, NADPH oxidation can be quantified by measuring the decrease in absorbance at this wavelength. A blank assay (solvent control,  $A_{\text{blank}}$ ) was run in the absence of catalysts. The rate of NADPH oxidation ( $v_0$ ) was calculated using a molar absorbance coefficient for NADPH of  $6.22 \times 10^3 \text{ M}^{-1} \text{ cm}^{-1}$ .

#### Activity of rabbit reticulocyte 12/15-LOX<sup>5</sup>

The enzyme (2  $\mu$ l of a 1.7 mg ml<sup>-1</sup> enzyme solution) was preincubated with the inhibitors (DMSO solution) at 22 °C in 0.1 M phosphate buffer (pH 7.4) for 5 min and the reaction was started by the addition of linoleic acid as substrate (20  $\mu$ l of a methanolic stock solution, 100  $\mu$ M final concentration, assay volume 1 ml). The catalytic activity of the enzyme was assayed spectrophotometrically following the increase in absorbance at 235 nm over a time interval of 30 s.

#### Activity of recombinant human 5-LOX<sup>29</sup>

The enzyme preparation (lysate) solution (10 µl) was incubated with the inhibitors (DMSO solution) for 10 min at room temperature in 0.1 M phosphate buffer (0.5 ml final volume, pH 7.4), containing 0.2 mM arachidonic acid, 0.4 mM CaCl<sub>2</sub>, 40µg mL<sup>-1</sup> dipalmitoyl phosphatidylcholine, 0.1 mM ATP and 0.1 mM EDTA. The oxygenated product (5-hydroperoxyeicosateraenoic acid) was then reduced with NaBH<sub>4</sub> (50 mg ml<sup>-1</sup>, 10 µl) to 5hydroxyeicosatetraenoic acid, the mixture was acidified with acetic acid (50 µl), 0.5 ml of ice-cold methanol was added and kept in ice for 10 min. 400 µl aliquot of the clear supernatant obtained by centrifuging (10000 rpm, 10 min) was injected to RP-HPLC (Shimadzu LC10 HPLC system connected to a SPD-M19AVP diode array detector, CC 250/4 Nucleosil 120-5 C18 column). A solvent system of MeOH-H<sub>2</sub>O-AcOH (80:20:0.1) was used at a flow rate of 1 ml min<sup>-1</sup>. The peak area in the time zone between 19-20 min corresponding to a 5-HETE fraction at 235 nm was compared with that of the control without inhibitor to calculate a relative inhibitory activity.

#### Iron-reducing activity<sup>30</sup>

The reduction of Fe(III) by test compounds was monitored using ferrozine, a reagent that avidly binds Fe(II), forming a complex with a very high extinction coefficient at 562 nm. The reaction mixture comprised 50 mM  $\gamma$ -morpholinopropanesulfonic acid (pH 2.0) buffer, 1 mM ferrozine (buffer solution), 50  $\mu$ M test compound (DMSO solution) and 100  $\mu$ M FeCl<sub>3</sub> (DMSO solution) in a final volume of 1 ml. The reaction was started by addition of FeCl<sub>3</sub> and increase of absorbance at 562 nm was monitored for 10 min. As control a mixture with DMSO instead of a test compound was used.

#### Acknowledgements

Financial support of the Alexander von Humboldt Foundation and the European Commission (LSHM-CT-2004-0050333) is acknowledged. The authors thank Drs M. Schlangen and R. Zeisberg, Profs L. Ernst and T. Braun, and Mmes Kaetel and Klose for recording spectra.

#### Notes and references

- 1 D. P. Gelain, R. J. S. Dalmolin, V. L. Belau, J. C. F. Moreira, F. Klamt and M. A. A. Castro, *Front. Biosci.*, 2009, 4457–4463.
- 2 J. Lu and A. Holmgren, J. Biol. Chem., 2009, 284, 723-727.
- 3 B. K. Sarma and G. Mugesh, Chem.-Eur. J., 2008, 14, 10603-10614.
- 4 G. Mugesh, W.-W. du Mont and H. Sies, *Chem. Rev.*, 2001, **101**, 2125–2179.
- 5 S.-C. Yu, A. Borchert, H. Kuhn and I. Ivanov, *Chem.–Eur. J.*, 2008, 14, 7066–7071.
- 6 G. Mugesh, A. Panda, H. B. Singh, N. S. Punekar and R. J. Butcher, *Chem. Commun.*, 1998, 2227–2228.
- 7 S. S. Zade, H. B. Singh and R. J. Butcher, *Angew. Chem., Int. Ed.*, 2004, **43**, 4513–4515.
- 8 R. E. Nelson, E. F. Degering and J. A. Bilderback, J. Am. Chem. Soc., 1938, 60, 1239–1241.
- 9 T. Schewe, Gen. Pharmacol., 1995, 26, 1153-1169.
- 10 H. Kuhn and V. B. O'Donnell, Prog. Lipid Res., 2006, 45, 334–356.
- 11 C. Berg, S. Hammarström, H. Herbertsson, E. Lindström, A. C. Svensson, M. Soederström, P. Tengvall and T. Bengtsson, *Thromb. Haemost.*, 2006, **96**, 652–659.
- 12 H. Kuhn, P. Chaitidis, J. Roffeis and M. Walther, J. Cardiovasc. Pharmacol., 2007, 50, 609–620.
- 13 B. S. Zweifel, M. M. Hardy, G. D. Anderson, D. R. Dufield, R. A. Pufahl and J. L. Masferrer, *Eur. J. Pharmacol.*, 2008, **584**, 166–174.
- 14 W. Berger, M. T. De Chandt and C. B. Cairns, Int. J. Clin. Pract., 2007, 61, 663–676.
- 15 B. Biteman, I. R. Hassan, E. Walker, A. J. Leedom, M. Dunn, F. Seta, M. Laniado-Schwartzman and K. Gronert, *FASEB J.*, 2007, 21, 2257– 2266.
- 16 G. Fürstenberger, N. Epp, K. M. Eckl, H. C. Hennies, C. Jørgensen, P. Hallenborg, K. Kristiansen and P. Krieg, *Prostaglandins Other Lipid Mediators*, 2007, 82, 128–134.
- 17 Complete crystallographic data (excluding structure factors) have been deposited at the Cambridge Crystallographic Data Centre under the numbers CCDC 740213 (**3c**), CCDC 740214 (**3d**) and CCDC 740215 (**6**).
- 18 R. K. Shirumalla, K. S. Naruganahalli, S. G. Dastidar, V. Sattigeri, G. Kaur, C. Deb, J. B. Gupta, M. Salman and A. Ray, *Inflammation Res.*, 2006, 55, 517–527.
- 19 L. Engman, D. Stern, H. Frisell, K. Vessman, M. Berglund, B. Ek and C. M. Andersson, *Bioorg. Med. Chem.*, 1995, 3, 1255–1262.
- 20 C. F. Lin, T. C. Chang, C. C. Chiang, J. Tsai and L. Y. Hsu, *Chem. Pharm. Bull.*, 2005, 53, 1402–1407.
- 21 D. Lapenna, G. Ciofani, S. D. Pierdomenico, M. Neri, C. Cuccurullo, M. A. Giamberardino and F. Cuccurullo, *Biochim. Biophys. Acta, Gen. Subj.*, 2009, **1790**, 25–30.
- 22 M. Walther, H.-G. Holzhutter, R. J. Kuban, R. Wiesner, J. Rathmann and H. Kuhn, *Mol. Pharmacol.*, 1999, 56, 196–203.
- 23 H. Kuhn, J. Saam, S. Eibach, H. G. Holzhütter, I. Ivanov and M. Walther, *Biochem. Biophys. Res. Commun.*, 2005, 338, 93–101.
- 24 K. Schwarz, M. Walther, M. Anton, C. Gerth, I. Feussner and H. Kuhn, J. Biol. Chem., 2001, 276, 773–779.
- 25 A. Bertelli, C. Armetti, A. Coco, G. Prino and P. Procopio, *Boll. Chim. Farm.*, 1967, **106**, 443–451.
- 26 R. E. Nelson, Proc. Ind. Acad. Sci., 1934, 44, 135-137.
- 27 S. Keimatsu, K. Yokota and I. Satoda, Yakugaku Zasshi, 1933, 53, 994–1046.
- 28 G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Crystallogr., 2008, 64, 112–122.
- 29 H. Kuhn, M. Anton, C. Gerth and A. Habenicht, *Arterioscler.*, *Thromb.*, *Vasc. Biol.*, 2003, 23, 1072–1076.
- 30 R. Dupont, J-F. Goossens, N. Cotelle, L. Vrielynck, H. Vezin, J.-P. Henichart and P. Cotelle, *Bioorg. Med. Chem.*, 2001, 9, 229–235.